

Effect of nasal air temperature on lung function

Authors' affiliations:

E. Millqvist, Asthma and Allergy Centre,
Sahlgrenska University Hospital, Gothenburg,
Sweden

Correspondence to:

Eva Millqvist MD
Asthma and Allergy Centre
Sahlgrenska University Hospital
S-413 45 Gothenburg
Sweden

Introduction

Inhalation of cold air is known to induce intrapulmonary airway obstruction in sensitive asthmatic patients and this is a condition often ascribed to heat and water losses from the airway mucosa (1–6). However, the pathophysiological mechanism explaining why airway cooling and exercise cause broncho-obstruction is not yet clear. The fact that anticholinergic drugs suppress exercise-induced asthma may indicate a reflex through the vagal nerve, although blockade of this nerve does not completely prevent broncho-obstruction induced by exercise in asthmatic patients (7–11). The fact that sodium cromoglycate prevents the development of exercise-induced asthma (9, 12–15) and that elevated levels of histamine are present in exercise-induced asthma (11, 16–18) indicate that mediators may be involved. The importance of the latter is uncertain, since antihistamines have a poor preventive effect in exercise-induced asthma (19). Changes in osmolarity of the airway lining may also be an important factor (20–22). Furthermore, asthmatic patients react with broncho-obstruction when challenged with hypo- and hyperosmolar solutions (23, 24) (Fig. 1).

Relation between upper and lower airways

Some evidence of a nasobronchial reflex between the upper and lower airways has been reported but its existence has not been established. In animal experiments, mechanical stimuli or insufflation of irritating chemical agents into the nose induces bronchodilation (25, 26) or broncho-obstruction (27, 28), or it may have no bronchial effect at all (29). In humans, mechanical or chemical stimuli in the nose and nasopharyngeal area induce increased resistance in

Date:

Accepted for publication 13 August 1998

To cite this article:

Millqvist E. Effect of nasal air temperature on lung function.

Allergy 1999, 54, Suppl 57, 106–111.

Copyright © Munksgaard 1999

ISSN 0105-4538

the intrapulmonary airways in healthy subjects (30). In the older German literature, a reflex from the nose to the lung via the trigeminus nerve was proposed – the “Wetter und Windreflex” (31).

According to the patients, obstruction is often rapidly induced by exposure to cold air. It seems unlikely that the degrees of heat and water losses from the airway mucosa provide the only explanation of these symptoms. Berger et al. noticed that cold stimulation in the nose of asthmatic patients increased the resistance of the lower airways (32). The effect could be blocked by first inhaling an anticholinergic drug. The authors believed that a reflex from the upper airways hastened the onset of obstruction. However, they did not prove that the cold air administered via the nose had failed to reach the intrapulmonary airways, nor did they use any controls. More recently, Fontanari et al. studied the consequences of nasal breathing of cold air in normal and asthmatic subjects and found an increase in interruption resistance that could be prevented by nasal anaesthesia or inhalation of a cholinergic antagonist (33, 34). None of these studies proved that the cold air did not reach the lower airways.

Cold air provocation at the nose and face, respectively, in healthy normals and asthmatic patients induced an increase in airway resistance (35–38). However, in these studies cold air may have reached the pulmonary airways and induced bronchial obstruction by cooling and/or drying of the airway

mucosa. Thus, although the existence of a nasobronchial reflex seems likely, at least following mechanical and chemical stimuli, it has not been demonstrated convincingly in connection with cold and warm stimuli.

The neural control of the human nose and lung is a complex system. The sensory innervation of the nose is derived mainly from the trigeminal nerve. In the human lung, parasympathetic efferent fibres are conducted by the vagus nerves to the parasympathetic ganglions in the bronchial wall. The baseline bronchomotor tone was thought initially to be regulated by a balance between contracting, parasympathetic impulses and dilating, adrenergic impulses having a direct influence on the smooth muscle cell via β -adrenoceptors. A third, efferent, non-adrenergic, noncholinergic pathway remains largely undefined, because no suitable inhibitor for this pathway or its neurotransmitter has yet been described (40–44).

In some individuals, cold air challenge of the nose leads to mast cell inflammatory mediator release (45–48) which could be a possible explanation to our results, described below.

From a clinical point of view, the relationship between asthma and sinusitis has often been discussed (49–51). In patients with allergic rhinitis, bronchial reactivity towards methacholine is increased (52) and in patients with perennial rhinitis, lower airway resistance increases after nasal challenge with histamine (53), but other studies have reported conflicting findings (54, 55). There is a well-known connection between nasal polyps, aspirin intolerance and asthma (56–58). In adults with asthma, intolerance to aspirin occur in 5–10% and in this group about half the patients have nasal polyps (58).

In the study presented below, we investigated whether changes in the nasal air temperature affected lung function in asthmatic patients and healthy subjects, when care was taken to prevent cold air from reaching the lower airways.

Patients and methods

Ten patients, seven women and three men (20–48 years) with a history of cold-sensitive asthma and eight healthy subjects, three women and five men (25–57 years), took part in this study. All patients had regular treatment with inhaled corticosteroids and used intermittent inhaled β_2 -stimulants. Two patients had regular treatment with inhaled long-acting β_2 -stimulants and one topical nasal corticosteroid. All patients had a history of atopy and had earlier shown an increase in FEV₁ of at least 20% after the

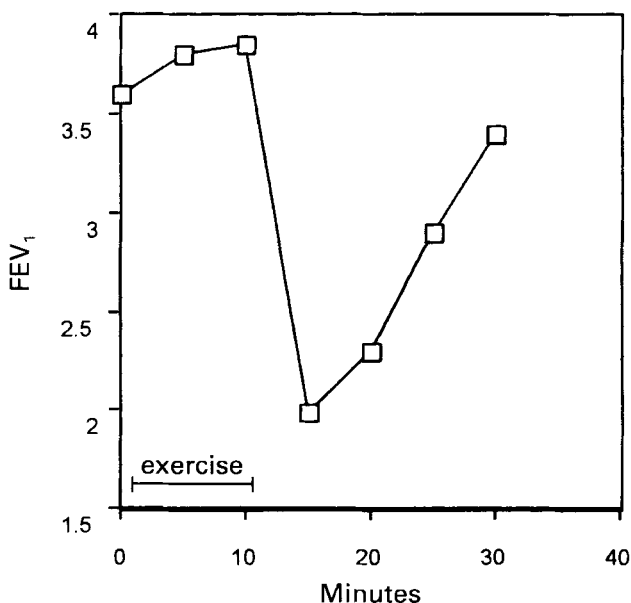


Figure 1. Typical course of exercise-induced asthma. FEV₁ increases during exercise, declines at the end of the exertion, with a maximum a few minutes after cessation, and then gradually returns to the initial level.

inhalation of a β_2 -stimulant. On most days they had no asthmatic symptoms. None of them smoked. No medication was taken for a minimum of 12 h before any test. Long-acting bronchodilators were withheld for at least 24 h.

The provocations were performed on 3 separate days, at approximately the same time each day, in a randomized order with air of various temperatures, i.e. cold air (about -15°C), ambient air (about $+22^\circ\text{C}$) or warm air (about $+37^\circ\text{C}$). At each nasal provocation, an airstream (0.6 l/s) was blown into both nostrils through a nose halter. A provocation consisted of 10 puffs of air of 15 s duration each at 1-min intervals between the puffs. In the halter, just at the edge of the nose, a thermistor recorded the temperature of the air. Cold air was obtained from an air cylinder, stored in a freezer. Ambient air and heated air were obtained from the central hospital system and were led through a temperate water-bath for moistening and regulation of the temperature. The airflow passing from the nose out through the mouth was monitored by a pneumotachograph connected to a mouthpiece. Shortly before each puff of air, the patients made a Valsalva manoeuvre. Positive intrathoracic pressure against the glottis was maintained during the provocation and for a few seconds thereafter. A constant positive pressure excluded any leakage of air from the nose to the lower airways during the nasal provocations. Pressure in the oesophagus was monitored by an oesophageal balloon, which was inserted through the mouth and swallowed without local anaesthesia. The pressure was recorded on a direct writer, together with the provocation air temperature and mouth flow.

Specific airway conductance (SGaw) and 1-s forced expiratory volumes (FEV₁) were determined before and 0, 5, 10 and 15 min after the provocations. SGaw was determined in a body plethysmograph and FEV₁ with a spirometer (Vitalograph, Buckingham, UK). Recordings of SGaw always preceded those of FEV₁ and the recordings were evaluated blindly.

Before each provocation an oesophageal balloon was swallowed. The patients then rested for a quarter of an hour before SGaw and FEV₁ were measured. Thereafter, the nasal provocation took place with the patient seated in an upright position and connected to the nose halter and the mouthpiece, as described above. After the 10 puffs of air lasting 15 s each at 1-min intervals (and therefore 10 Valsalva manoeuvres), the balloon in the oesophagus was removed and again the symptom scores, heart rate and blood pressure were recorded. This was again followed by recording SGaw and FEV₁.

The effects of these provocations were analysed by calculating "the area under the curve" – i.e. an expression of the overall difference from baseline and by analysis of variance (ANOVA) and by the Wilcoxon signed rank test. *P*-values less than 0.05 were considered significant.

Results

Changes in SGaw in percent from baseline after the three provocations are illustrated in Fig. 2. Cold provocation caused a significant fall of SGaw and warm air provocation a significant rise in the asthmatic patients (left panel), whereas there were no significant effects of room air provocations. Furthermore, the differences in SGaw-areas under the curves were significantly different between all three provocations. Ten minutes after the cold air provocation SGaw fell, on average, by 23% ($P < 0.005$) and rose directly after the provocation with warm air on average 15% ($P < 0.05$). In the normal subjects (right panel) there were no significant differences.

Fig. 3 shows the corresponding results for FEV₁. In the asthmatics (left panel) FEV₁ fell on average by 8% ($P < 0.05$) 5 min after the cold air provocation and rose directly after warm air provocation on average by 6% ($P < 0.05$). Analysis of the area under the curve revealed that all three provocations differed significantly in the asthmatic patients, in accordance with the SGaw recordings. In the healthy subjects (right panel) the provocations caused no significant differences.

Discussion

The present results confirm the existence of an interplay between the nose and the rest of the airways; the mechanism could be due to a trigeminal reflex or a mediator release.

Cold air stimulation at the nose induced a decrease in airway conductance and FEV₁, and warm air stimulation had the opposite effect. Although it is believed that the nasal mucosa lacks temperature receptors, it has been shown that hyperosmolar stimuli can induce naso-nasal reflexes (47). These sensory nerves could possibly signal to the central nervous system on the basis of the osmolarity of the intraepithelial junction fluid. This continuous impulsive may result in nasal glandular secretion and baseline smooth muscle tone being regulated by the nasal mucosa. When warm, moist air is inhaled the osmolarity is decreased

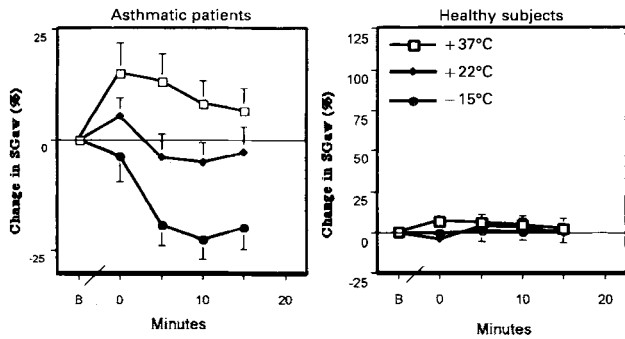


Figure 2. Changes in SGaw after nasal provocations with -15°C , $+22^{\circ}\text{C}$ and $+37^{\circ}\text{C}$. The results are shown as mean values expressed in percentage of the preprovocation values. Mean values and SE are given. "B" denotes before provocation.

and the impulse rate may drop precipitously. This would result in reduction in airway tone followed by bronchodilation.

We do not know the reason for the discrepancy in the effects of nasal air temperature between asthmatics and normals. Rhinitis is characterized by mucosal hyperreactivity and found in almost all asthmatic patients. Hyperreactivity is, to a great extent, neuronal in nature (subjects with perennial rhinitis respond more intensely to capsaicin and to cold dry air than healthy individuals (60, 61)). It is possible that cold air generate a central spasmogenic reflex by stimulating hyperreactive sensory nerve endings and that this only occurs in a hyperreactive, not healthy nose. Another possibility is that the signal is generated by the nasal mucosa of both groups but leads to bronchial obstruction only in asthmatics. Perhaps the reactions are less pronounced in normal people and therefore remained undetected in the present limited study.

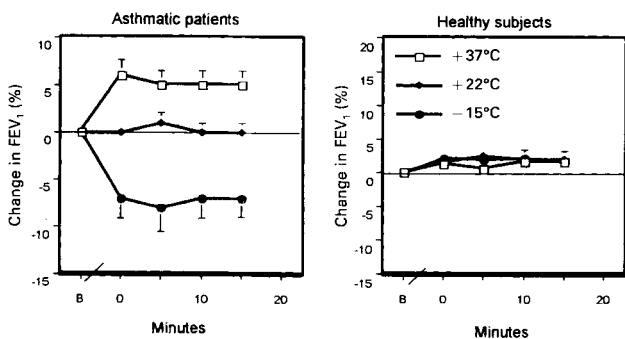


Figure 3. Changes in FEV₁ after nasal provocations with -15°C , $+22^{\circ}\text{C}$ and $+37^{\circ}\text{C}$. The results are shown as mean values expressed as percentage of the pre-provocation values. Mean values and SE are given. "B" denotes before provocation.

The changes in airway conductance and FEV₁ after cold or warm air provocation may have been due to varying airway muscle tone, to varying degrees of oedema of the airway mucosa, to varying amounts of mucus secretions in the airway lumen or to any combination of these factors. The changes occurred immediately after the provocations, a fact that may indicate a change in muscle tone as the major mechanism. After the provocation with cold air the depression of lung function seemed to occur quicker when measured with SGaw compared to FEV₁. This might be dependent on the SGaw measurements always preceding those of FEV₁ (and thus there was some minutes delay for the FEV₁ measurements) or the possibility that these methods reflected different parts of the lungs, e.g. small and large airways. The results of SGaw and FEV₁ after the provocations are consistent, although there were some individual variations.

During nasal breathing at rest, the inspired air temperature at the carina is about 36°C when breathing room air (about 20°C) and about 33°C when breathing air of -20°C . The inspired air in the trachea is almost saturated with water, irrespective of inspired humidity and temperature. The corresponding air temperatures at the carina during oral breathing are 35°C and 32°C at rest, but there is a fall to about 25°C during maximal hyperventilation with cold air (62, 63). When asthmatic patients hyperventilated cold air, their bronchial obstruction was found to be less severe during nasal than oral breathing (6). If breathing cold air through the nose activates a nasobronchial reflex, its effect is offset by the conditioning capability of the nose. During exercise, however, most people switch to oronasal breathing (64) and bypass much of the heat and moisture exchanging properties of the nose.

Conclusions

This study confirms a relationship between the upper and lower airways. In asthmatic patients, cold air administered in the nose caused broncho-obstruction and warm air resulted in bronchodilation. What may be the importance of a nasobronchial interplay? This appears to be of little or no importance in healthy subjects. In asthmatic patients it may act as a warning signal to prevent the airways from additional exposure to cold air. It is a clinical experience that some asthmatic patients have a very rapid onset of cold-induced asthma symptoms and this could originate from the nose. Further knowledge of the physiology related to the nasobronchial interplay may be of therapeutic value in treating asthma.

References

1. Anderson SD, Schoeffel RE, Follet R, Perry CP, Daviskas E, Kendall M. Sensitivity to heat and water loss at rest and during exercise in asthmatic patients. *Eur J Respir Dis* 1982;63:459-471.
2. Chen WY, Horton DJ. Heat and water loss from the airways and exercise-induced asthma. *Respiration* 1977;34:305-323.
3. Deal EC, McFadden ER, Ingram RH, Strauss RH, Jaeger JJ. Role of respiratory heat exchange in production of exercise-induced asthma. *J Appl Physiol* 1979;46:467-475.
4. Strauss RH, McFadden ER, Ingram RH, Deal EC, Jaeger JJ. Influence of heat and humidity on the airway obstruction induced by exercise. *J Clin Invest* 1978;61:433-440.
5. Strauss RH, McFadden ER, Ingram RH, Jaeger JJ. Enhancement of exercise-induced asthma by cold air. *N Engl J Med* 1977;297:743-747.
6. Griffin MP, McFadden JR, Ingram RH. Airway cooling in asthmatic and nonasthmatic subjects during nasal and oral breathing. *J Allergy Clin Immunol* 1982;69:354-359.
7. Poppius H, Salorinne Y. Comparative trial of salbutamol and an anticholinergic drug, SCH 1000, in prevention of exercise-induced asthma. *Scand J Respir Dis* 1973;54:142-147.
8. Godfrey S, König P. Inhibition of exercise-induced asthma by different pharmacological pathways. *Thorax* 1976;31:137-143.
9. Anderson S, Seale JP, Ferris L, Schoeffel R, Lindsay DA. An evaluation of pharmacotherapy for exercise-induced asthma. *J Allergy Clin Immunol* 1979;64:612-624.
10. Breslin FJ, McFadden ER, Ingram RH, Chandler D. Effects of atropine on respiratory heat loss in asthma. *J Appl Physiol* 1980;48:619-623.
11. Finnerty JP, Holgate ST. The contribution of histamine release and vagal reflexes, alone and in combination, to exercise-induced asthma. *Eur Respir J* 1993;6:1132-1137.
12. Fanta CH, McFadden ER, Ingram RH. Effects of cromolyn sodium on the response to respiratory heat loss in normal subjects. *Am Rev Respir Dis* 1981;123:161-164.
13. Breslin FJ, McFadden ERJ, Ingram RHJ. The effects of cromolyn sodium on the airway response to hyperpnea and cold air in asthma. *Am Rev Respir Dis* 1980;122:11-16.
14. Morton AR, Turner KJ, Fitch KD. Protection from exercise-induced asthma by pre-exercise cromolyn sodium and its relationship to serum IgE levels. *Ann Allergy* 1973;31:265-271.
15. Wallace D, Grieco MH. Double-blind, cross-over study of sodium inhibition of exercise-induced bronchospasm in adults. *Ann Allergy* 1976;37:153-163.
16. Barnes PJ, Brown MJ. Venous plasma histamine in exercise- and hyperventilation-induced asthma in man. *Clin Sci* 1981;61:159-162.
17. Lee TH, Brown MJ, Nagy L, Clauson R, Walport MJ, Kay AB. Exercise-induced release of histamine and neutrophil chemotactic factor in atopic asthmatics. *J Allergy Clin Immunol* 1982;70:73-81.
18. Belcher NG, Murdoch R, Dalton N, Clark TJH, Rees PJ, Lee TH. Circulating concentrations of histamine, neutrophil chemotactic activity, and catecholamines during the refractory period in exercise-induced asthma. *J Allergy Clin Immunol* 1988;81:100-110.
19. Zielinski J, Chodosowska E. Exercise-induced bronchoconstriction in patients with bronchial asthma. Its prevention with an antihistaminic agent. *Respiration* 1977;34:31-35.
20. Lilker ES, Jauregui R. Airway response to water inhalation: a new test for "bronchial reactivity". *N Engl J Med* 1981;305:702.
21. Anderson SD, Schoeffel RE, Black JL, Daviskas E. Airway cooling as the stimulus to exercise-induced asthma - a re-evaluation. *Eur J Respir Dis* 1985;67:20-30.
22. Dejaegher P, Rochette F, Clarysse I, Demedts M. Hypocapnic hyperventilation versus isocapnic hyperventilation with ambient air or with dry air in asthmatics. *Eur J Respir Dis* 1987;69:102-109.
23. Schoeffel RE, Anderson SD, Altounyan ECA. Bronchial hyperreactivity in response to inhalation of ultrasonically nebulised solutions of distilled water and saline. *Br Med J* 1981;288:1285-1287.
24. Makker HK, Holgate ST. Relation of the hypertonic saline responsiveness of the airways to exercise induced asthma, symptom severity, and to histamine or methacholine reactivity. *Thorax* 1993;48:142-147.
25. Tomori Z, Widdicombe JG. Muscular bronchomotor and cardiovascular reflexes elicited by mechanical stimulation of the respiratory tract. *J Physiol* 1968;200:25-49.
26. Allison DJ, Clay TP, Hughes JMB, Jones HA, Shevis A. Effects of nasal stimulation on total respiratory resistance in the rabbit. *Physiol Soc* 1974;239:23-24.
27. Ellis M. The mechanism of the bronchial movements and the naso-pulmonary reflex. *Proc R Soc Med* 1936;29:527-536.
28. Angell-James JE, Burgh D, Daly M. Some aspects of the upper respiratory tract reflexes. *Acta Otolaryngol (Stockh)* 1975;79:242-252.
29. Nadel JA, Widdicombe JG. Reflex effects of upper airway irritation on total lung resistance and blood pressure. *J Appl Physiol* 1962;17:861-865.
30. Kaufman J, Wright GW. The effect of nasal and nasopharyngeal irritation on airway resistance in man. *Am Rev Respir Dis* 1969;100:626-630.
31. Sercer A. Über die Beeinflussung der Bronchien von der Nase aus. *Arch Ohr Nas Kehlkopf Heilk* 1952;161:264-275.
32. Berger D, Nolte D. On nasobronchial reflex in asthmatic patients. *Rhinology* 1979;17:193-198.
33. Fontanari P, Burnet H, Zattara-Hartmann M, James Y. Changes in airway resistance induced by nasal inhalation of cold dry, dry, or moist air in normal individuals. *J Appl Physiol* 1996;81:1739-1743.
34. Fontanari P, Zattara-Hartmann M, Burnet H, James Y. Nasal eucapnic inhalation of cold, dry air increases airway resistance in asthmatic patients. *Eur Respir J* 1997;10:2250-2254.
35. Josenhans WT, Melville GN, Ulmer WT. The effect of facial cold stimulation on airway conductance in healthy man. *Can J Physiol Pharmacol* 1969;47:453-457.
36. Melville NG, Morris D. Effect on airway resistance in health and disease. *Environ Physiol Biochem* 1972;2:107-116.
37. Berk JL, Lenner KA, McFadden ER. Cold-induced bronchoconstriction: role of cutaneous reflexes vs. direct airway effects. *J Appl Physiol* 1987;63:659-664.
38. Koskela H, Tukiainen H. Facial cooling, but not nasal breathing of cold air, induces bronchoconstriction: a study in asthmatic and healthy subjects. *Eur Respir J* 1995;8:2088-2093.
39. McDonald J, Nelson J, Lenner K, McLane M, McFadden EJ. Effects of the combination of skin cooling and hyperpnea of frigid air in asthmatic and normal subjects. *J Appl Physiol* 1997;82:453-459.
40. Richardson JB, Beland J. Nonadrenergic inhibitory nervous system in human airways. *J Appl Physiol* 1976;41:764-771.
41. Richardson JB. Nerve supply to the lungs. *Am Rev Respir Dis* 1979;119:785-802.
42. Richardson JB. Recent progress in pulmonary innervation. *Am Rev Respir Dis* 1983;128:65-68.
43. Barnes PJ, Baraniuk JN, Belvisi MG. Neuropeptides in the respiratory tract. *Am Rev Respir Dis* 1991;144:1187-1198.
44. Baraniuk JN. Neural control of human nasal secretion. *Pulmon Pharmacol* 1991;4:1-31.

45. Togias AG, Naclerio RM, Proud D. Nasal challenge with cold, dry air results in the production of inflammatory mediators: possible mast cells involvement. *J Clin Invest* 1985;76:1375-1381.
46. Togias A, Naclerio R, Proud D, et al. Studies on the allergic and nonallergic inflammation. *J Allergy Clin Immunol* 1988;81:782-790.
47. Kraysenbuhl M, Hudspeth B, Brostoff J, Scadding G, Guesdon J, Lachman Y. Nasal histamine release following hyperosmolar and allergen challenge. *Eur J Allergy Clin Immunol* 1989;44:25-29.
48. Proud D, Baily GS, Naclerio RM. Tryptase and histamine as markers to evaluate mast cell activation during the responses to nasal challenge with allergen, cold dry air and hyperosmolar solutions. *J Allergy Clin Immunol* 1992;89:1098-1110.
49. Slavin RG. Sinusitis in adults and its relation to allergic rhinitis, asthma and nasal polyps. *J Allergy Clin Immunol* 1988;82:950-956.
50. Brown JA. Sino-bronchial reflex, asthma and sinusitis. *J S Carol Med Assoc* 1992;88:340-343.
51. Slavin RG. Sinopulmonary relationships. *Am J Otolaryngol* 1994;15:18-25.
52. Ramsdal EH, Morris MM, Robert RS, Hargreave FE. A symptomatic bronchial hyperresponsiveness in rhinitis. *J Allergy Clin Immunol* 1985;75:573-577.
53. Yan K, Salome C. The response of the airways to nasal stimulation in asthmatics with rhinitis. *Eur J Respir Dis* 1983;64(Suppl. 128):105-108.
54. Small P, Biskin N. The effects of allergen-induced nasal provocation on pulmonary function in patients with perennial allergic rhinitis. *Am J Rhinol* 1989;3:17.
55. Litell NT, Carlisle CC, Millman RP, et al. Changes in airway resistance following nasal provocation. *Am Rev Respir Dis* 1990;141:580.
56. Delaney J. Aspirin idiosyncrasy in patients admitted for nasal polypectomy. *Clin Otolaryngol* 1976;1:27-30.
57. Stenius BSM, Lemola H. Hypersensitivity to acetylsalicylic acid (ASA) and tartrazine in patients with asthma. *Clin Allergy* 1976;6:119.
58. Szczeklik A. Analgesics, allergy and asthma. *Br J Clin Pharmacol* 1980;10:401-404.
59. Millqvist E, Bende M, Johansson Å, Bake B. Effect of nasal air temperature on lung function. Submitted 1999.
60. Philip G, Baroody F, Proud D, Naclerio R, Togias A. Increased responses to capsaicin in nasal challenge in allergic versus non-allergic rhinitis. *J Allergy Clin Immunol* 1994;94:1035-1045.
61. Philip G, Jankowski R, Baroody F, Naclerio R. Reflex activation of nasal secretion by unilateral inhalation of cold dry air. *Am Rev Respir Dis* 1993;148:1616-1622.
62. Cole P. Recordings of respiratory air temperature. *J Laryngol* 1954;68:295-307.
63. McFadden ER, Pichurko BM, Bowman HF, et al. Thermal mappings of the airways in humans. *J Appl Physiol* 1985;58:564-570.
64. Niinmaa V, Cole P, Mintz S, Shepard RJ. The switching point from nasal to oronasal breathing. *Respir Physiol* 1980;42:61-71.